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(54) **Repair of tissue in animals.**

(57) A composition for the promotion of cell proliferation and tissue repair in animals having as active ingredients a TGF- β which is activated by either a TGF- α or an EGF or both; and methods for administration. As another embodiment these active ingredients can be admixed with other (secondary) growth factors.

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REPAIR OF TISSUE IN ANIMALS

This is a continuation-in-part application of USSN 06/468,590, filed 22 February 1983 (now pending), which is in turn a continuation-in-part application of USSN 06/423,203, filed 24 September 1982 (now abandoned).

BACKGROUND OF THE INVENTION1. Field Of The Invention

5 This invention relates to compositions which promote repair of tissue, particularly fibroblast cells, in animals, particularly human beings. This invention further relates to a method of treating wounds by the topical or systemic administration of the compositions.

2. Description Of The Prior Art

10 There is a continuing need for the promotion of rapid cell proliferation at the site of wounds, burns, diabetic and decubitus ulcers, and other traumata.

15 A number of "growth factors" are known, which promote the rapid growth of animal cells. These growth factors include epidermal growth factor (EGF), transforming growth factors (TGF's), and nerve growth factor (NGF). However, prior to this invention, none of these growth factors have been found to be
20 pharmaceutically acceptable agents for the acceleration of wound healing.

It has been shown that the mitogenic activity of insulin (a hormone) can be increased many-fold by the presence of prostaglandin $F_{2\alpha}$ (not exactly a hormone, but having similar properties - it causes constriction of vascular smooth muscle), see L. Jimenez de Asua, et al., Cold Spring Harbor Conf. Cell Proliferation Vol. 6, Sato, ed., Cold Spring Harbor Labs., New York, (1979) at Pp. 403-424. Similar activation of insulin has been reported with fibroblast growth factor by P. S. Rudland, et al., Proc. Natl. Acad. Sci., U.S.A., 71:2600-2604 (1974) and with EGF by R. W. Holley, et al., Proc. Natl. Acad. Sci., U.S.A., 71:2908-2911 (1974). Furthermore, in the "competance-progression" scheme of C. D. Stiles, et al., in Proc. Natl. Acad. Sci., U.S.A., 76:1279-1283 (1979), positive effects on cell growth have been demonstrated for platelet-derived growth factor or fibroblast growth factor in combination with members of the insulin family such as somatomedins A and C, the insulin-like growth factors.

Many new peptide growth factors have been isolated and characterized recently, as indicated in Tissue Growth Factors, R. Baserga, ed., Springer-Verlag pub., New York (1981), however there have been few studies on the activity of these materials in vivo. In many cases, the relatively small amounts of peptides available have limited the ability to study their properties in vivo. An important area for potential application of peptide growth factors is the enhancement of wound healing. Despite the need for rapid wound healing in the treatment of severe burns, trauma, diabetic and

12. The method of Claim 11, further comprising periodic topical reapplication.

13. The method of Claim 11 further comprising continuing topical application.

14. A method of treating mammalian systemic traumata comprising at least one systemic administration of the composition of Claim 10 in a pharmaceutically acceptable carrier.

15. The method of Claim 14, further comprising periodic systemic readministration.

16. The method of Claim 14, further comprising continual systemic administration.

17. The composition of claim 1 wherein the activated TGF- β is admixed with at least one secondary growth factor in a mole ratio of 1:.1-10.

18. The composition of claim 17 wherein the secondary growth factor is at least one the group consisting of: platelet-derived growth factors, fibroblast growth factors, angiogenesis growth factors, insulin-like growth factors, insulin, nerve growth factors, and anabolic steroids.

19. The composition of claim 17 wherein the secondary growth factor is at least one of the group consisting of platelet-derived growth factors, fibroblast growth factors and angiogenesis factors.

20. The composition of claim 18 wherein the activated TGF- β and secondary growth factors are present in about equimolar amounts.

21. The method of claim 11 wherein the activated TGF- β is admixed with at least one secondary growth factor in a mole ratio of 1:.1-10.

22. The method of claim 21 wherein the secondary growth factor is at least one of the group consisting of: platelet-derived growth factors, fibroblast growth factors, angiogenesis growth factors, insulin-like growth factors, insulin, nerve growth factors, and anabolic steroids.

23. Beta-type transforming growth factor.

24. The growth factor of claim 23 derived from a mammalian source.

25. The growth factor of claim 23 derived from a genetically engineered cell.

26. The growth factor of claim 24 derived from a bovine kidney and purified.

27. The growth factor of claim 24 derived from a human source.

activity in vivo is necessarily related to the development of malignancy. Indeed, the transformed phenotype is a physiological state associated with normal embryological development, and transforming (onc) genes have been found in normal cells of essentially all vertebrates. The function of these onc genes from normal cells is not known at present. While there may be irreversible and excessive expression of TGF activity during malignant cell growth, the data at hand indicate that TGFs have a more benign and perhaps essential role in the function of normal cells. At present, it is not known what the intrinsic physiological roles of TGFs are. In this respect, TGFs are like many other peptide hormones or hormone-like agents which have recently been discovered and isolated; this is particularly true for many peptides of the nervous system, for which a defined chemical structure may be known, yet whose physiology is still a matter of uncertainty.

The initial description of sarcoma growth factor (SGF), the first of the TGFs to be isolated, was an important finding in tumor cell biology since it provided a direct mechanism for the expression of the neoplastic phenotype in a virally-transformed cell. Two important properties of SGF were described in these earliest studies, namely (1) that the effects of SGF in causing phenotypic transformation were dependent on its continued presence, and that these effects were reversible when SGF was removed, and (2) that the effects of SGF could be expressed in the very same cells that synthesized this peptide, a property that has been termed autocrine secretion. Although these two properties have not been definitively shown for all of the other more newly discovered TGFs, the functions of the entire set of TGFs can most reasonably

be assumed to be that of local, hormone-like agents that reversibly control cell function by paracrine or autocrine mechanisms.

5 Since the discovery of SGF in 1978, many TGFs
have been described from diverse sources. These
TGFs can be categorized into two groups: extracellular
TGFs isolated from conditioned media of cultured cells,
and intracellular (cell-associated) TGFs isolated by
direct extraction of cells or tissues. Although
10 extracellular TGFs have recently been isolated from
non-neoplastic murine cells, use of conditioned medium
has, in general, been restricted to neoplastic cell
lines that could be grown in long-term, large-scale
culture, including certain virally and chemically-
15 transformed rodent cells and human tumor cell lines.
The adaptation of an acid/ethanol extraction procedure
to TGF isolation removed all limitations on cell types
and quantities of tissues that could be examined.
Using this procedure, extracts of all tissues or cells,
20 whether of neoplastic or non-neoplastic origin; whether
from adult or embryonic tissue, whether from human,
bovine, or murine genomes, have been shown to promote
colony formation in a soft agar assay; hence, by
definition, these extracts contain TGF activity.

25 A variety of both epithelial and fibroblastic
cell lines form colonies in soft agar in the presence
of TGFs. However, the most commonly used indicator
cell line is the rat kidney fibroblast cell clone,
NRK 49F, which has been selected for its strong colony-
30 forming response to the TGFs. Rat-1 cells and mouse
AKR-2B cells have also been used successfully as
indicator cells.

5 All of the TGFs referred to above are low
molecular weight polypeptides which share with SGF the
physical properties of acid and heat stability and
sensitivity to treatment with both trypsin and
10 dithiothreitol. However, there are marked differences
in the biological properties of these TGFs, particu-
larly with respect to their relationship to EGF.
Certain TGFs, though antigenically distinct from EGF,
have some structural homology to EGF, since they
15 compete with EGF for receptor binding. Other TGFs do
not compete with EGF for receptor binding, but instead
are dependent on EGF for activity in the soft agar
assay for colony formation. To remove the ambiguities
implicit in the assignment of the general term "TGF"
20 to these different factors, an operational classifica-
tion of the members of the TGF family based on their
interactions with EGF is suggested, both with respect
to competition for binding to the EGF receptor and to
the requirement for EGF for induction of colonies in
soft agar.

As defined for purposes of this invention,
TGF- α are those TGFs which compete with EGF for receptor
binding and which do not require EGF for the induction
of colony formation in soft agar. TGFs with these
25 properties include SGF and other TGFs derived from
neoplastic cells, as well as some TGFs from mouse
embryos.

As defined for purposes of this invention,
TGF- β are those TGFs which do not compete with EGF for
30 receptor binding and which require EGF for the induction
of colony growth in soft agar. When assayed in the
presence of EGF, TGF- β represents the major colony-
forming activity of the intracellular TGFs of both neo-
plastic and non-neoplastic cell lines and tissues. It

can be assumed that TGF- β will be found in conditioned media as well, once the proper assays are used.

Those TGFs which do not compete with EGF for receptor binding and which do not require EGF for colony formation are designated TGF- γ (gamma-type TGF). Such TGFs have been described in conditioned media of certain virally or chemically transformed cells. Finally, TGF- δ (delta-type TGF) is used to specify those TGFs which would both compete for EGF receptors and require EGF for colony formation in soft agar. EGF itself could be classified as a weak TGF- δ .

Example Of Purification And Properties

Research in our laboratory has been directed toward the isolation of TGFs directly from cells and tissues. An acid/ethanol extraction procedure was modified for this purpose, as disclosed in A. B. Roberts, et al., Proc. Natl. Acad. Sci., U.S.A., 77:3494-3498 (1980), and chromatography and high pressure liquid chromatography (HPLC) have been employed for further purification. TGF activity, measured by the ability to induce non-neoplastic indicator cells (NRK) to form colonies in soft agar, has been quantitated on an image analysis system with respect to both the number and size of the colonies formed. By use of HPLC, we have shown that two distinctly different TGFs, here classified as TGF- α and TGF- β , can be isolated from the same pool of acid/ethanol extracts of MSV-transformed 3T3 cells; for this purpose, columns

using a linear gradient of acetonitrile in 0.1% trifluoroacetic acid have been used. TGF- α , which elutes from the column earlier than marker EGF, is characterized by its ability to induce the formation of small colonies (850-3,100 μm^2) in soft agar in the absence of added EGF and its ability to compete with EGF in a radio-receptor assay. TGF- β , which elutes later than TGF- α or marker EGF, does not compete with EGF for receptor binding and requires EGF to induce the formation of large colonies ($>3,100 \mu\text{m}^2$) in the soft agar assay.

TGF- α from MSV-transformed 3T3 cells resembles SGF isolated from the conditioned medium of the same cells and other TGFs isolated from rat and human tumor cell lines. Recently, SGF and the TGF- α 's from the conditioned media of a human melanoma cell line and virally-transformed rat embryo fibroblasts have been purified to homogeneity. The human melanoma TGF- α is a single chain polypeptide of molecular weight 7,400. Its amino acid composition and chromatographic behavior are markedly different from that of human EGF, but similar to that of murine SGF and rat TGF- α , suggesting that TGF- α 's from human, rat and mouse genomes are more closely related to each other than to EGF. There is therefore a reasonable possibility that TGF- α 's may have cross-species utility.

In sarcoma virus-transformed rodent cell lines, the release of TGF- α into the medium has been correlated with the expression of the transformed phenotype, and within a selected set of human tumor cell lines that release TGF- α , the ability of the tumor cells to grow in soft agar has been correlated with the quantity of TGF- α they secrete. However, secretion of TGF- α is not an absolute requirement for neoplastic behavior; certain

chemically transformed murine cell lines and human lung carcinoma cell lines that do not secrete TGF- α release strong TGF activity that does not compete with EGF for receptor binding.

5 TGF- β of the acid/ethanol extract of the MSV-transformed 3T3 cells resembles other TGFs isolated from many neoplastic and non-neoplastic tissues. After further purification on a second HPLC column, TGF- β of the MSV-transformed cells eluted at the
10 same position in the n-propanol gradient (48%) as one peak of TGF- β activity of the bovine salivary gland, and each was associated with a small peak of absorbance at 280 nm. These two TGF- β 's, one
15 from a neoplastic mouse cell line and the other from a non-neoplastic bovine tissue, each migrated as a 12,500-13,000 daltons MW protein on SDS-PAGE in the presence of mercaptoethanol and as an apparent 25,000-26,000 daltons protein in the absence of
20 mercaptoethanol; they therefore appear to be closely related to each other and different from both TGF- α and EGF. The finding of TGF- β in all non-neoplastic tissues examined thus far suggests a normal physiological function for these TGFs. There
25 is therefore a reasonable possibility that TGF- β 's may have cross-species utility.

Amino Acid Composition and Sequencing of TGF- β

Through a combination of techniques, TGF- β from bovine kidneys was purified 200,000-fold to the point of homogeneity.

Samples of homogenous TGF- β (20-50 pmol) were taken to dryness and hydrolyzed in sealed, evacuated tubes at 150°C for 2 hours in 100 μ l constant boiling HCl containing 0.1% liquid phenol.

5 Half-cystine and methionine were determined by performic acid oxidation followed by acid hydrolysis. Analyses of o-phthalaldehyde derivatives of the amino acids were done on a modified amino acid analyzer equipped with a fluorometer and computing

10 integrator.

For amino-terminal sequence analysis, approximately 500 picomoles (M_r 25,000) of TGF- β were reduced and S-carboxymethylated with dithiothreitol and iodo-[14 C] acetic acid in the presence

15 of 6M guanidine-HCl in 1 M Tris-HCl buffer, pH 8.4. Excess reagents were separated from carboxymethylated protein by HPLC on a 5 micron 50 x 4.6 mm column eluted with a gradient of 0-90% acetonitrile (1% per min) in 0.1% TFA. Overall recovery of the

20 procedure was 96%, based on estimating the amount of protein by amino acid analysis using fluorescamine detection.

Automated Edman degradation was performed on about 500 pmoles (M_r 12,500) of the S-carboxymethylated

25 protein with a gas-phase sequencer. PTH-amino acids were identified using an HPLC system. Initial yeild was about 30% and repetitive yield about 90%.

The following table summarizes the amino acid distribution.

TABLE 1

Amino Acid Composition of Bovine Kidney TGF- β (a)

<u>Amino Acid</u>	<u>Residues/mole</u> ^(b) (mean \pm range)
Aspartic acid	25 \pm 1
Threonine	7 \pm 1
Serine	16 \pm 2
Glutamic acid	26 \pm 1
Proline	N.D.
Glycine	15 \pm 3
Alanine	18 \pm 1
Half-cystine ^(c)	16 \pm 2
Valine	17 \pm 1
Methionine ^(c)	3 \pm 0
Isoleucine	11 \pm 1
Leucine	25 \pm 1
Tyrosine	17 \pm 2
Phenylalanine	8 \pm 1
Histidine	8 \pm 1
Lysine	22 \pm 2
Tryptophan	N.D.
Arginine	12 \pm 1

^aAmino acid composition of purified bovine kidney type β TGF determined after 2 h hydrolysis in 6 N HCl at 150°. Values are based on 3 separate determinations of 3 different preparations.

5 ^bThe number of residues per mole for each amino acid is based on an apparent molecular weight of 25,000.

^cDetermined by performic acid oxidation and acid hydrolysis.

N.D. - not determined

10 Analysis of the bovine kidney TGF- β by
electrophoresis on NaDodSO₄-polyacrylamide gels
suggests that some of the disulfide bonds are inter-
chain. Comparison of the above results with those
15 obtained by the analysis of TGF- β from human placenta
or human platelets revealed no significant differences
in the respective amino acid compositions, and it is
anticipated that the N-terminal sequences will be
very similar.

20 Amino acid sequence analysis of the reduced
and S-carboxymethylated bovine kidney TGF- β by
automated Edman degradation using a gas-phase
sequencer revealed a single N-terminal amino acid
sequence as follows, (CMC is S-carboxymethylcysteine):

Ala-Leu-Asp-Thr-⁵Asn-Tyr-CMC-Phe-Ser-¹⁰Ser-Thr-Glu-Lys-¹⁵Asn-CMC-.

Initial and repetitive yields were found to be equal to the yields calculated for myoglobin used as standard protein. At the minimum, the results indicate that the sequence of at least the

5 first fifteen N-terminal amino acids of each of the two subunits of TGF- β is identical and confirm the observations of a single protein band of the reduced TGF- β on NaDodSO₄-polyacrylamide gels. In addition, the N-terminal sequence of the bovine

10 kidney TGF- β is identical to the partial sequence of TGF- β from human placenta, suggesting a high degree of relatedness of type β TGFs from different species and different tissue sources.

Activation of TGF- β

15 Recent experiments in our laboratory have shown that either TGF- α or EGF will activate TGF- β to induce the formation of large colonies in soft agar. Purified

5 TGF- β from the MSV-transformed 3T3 cells, assayed by
itself, had no colony-forming activity at concentrations
as high as 2 $\mu\text{g/ml}$. However, assayed after activation
by the presence of either EGF, or TGF- α derived from the
10 same cells, TGF- β induced a dose-dependent formation of
large colonies ($>3,100 \mu\text{m}^2$) at concentrations of 10-200
ng/ml. By contrast, EGF or TGF- α , assayed by themselves,
induced a maximal response of only a small number of
colonies; this response was increased 10-fold by the
15 addition of TGF- β . The relative abilities of EGF
and TGF- α to promote TGF- β -dependent formation of large
colonies in soft agar correlated with their relative
abilities to compete for binding to the EGF receptor;
other experiments using chemically-modified EGF ana-
logues have substantiated this correlation. These
20 data, demonstrating that the induction of a strong
colony-forming response requires both TGF- α and TGF- β
or EGF, suggest that TGF- β , which is found in all
tissues, may be an essential mediator of the effects
of TGF- α and of EGF on neoplastic transformation.

Little is known about the mechanisms by which
exogenous TGFs induce non-neoplastic cells to express
the transformed phenotype. Furthermore, the synergistic
25 interactions of TGF- α and TGF- β suggest that these two
TGFs may act through different pathways. Experiments
using TGFs of conditioned media of sarcoma virus-
transformed rodent cells have shown that the synthesis
of new RNA and protein is required before transformation
occurs. Other experiments have been directed at a
30 possible role of TGFs in phosphorylation reactions.
Certain viral transforming gene products and their
normal cellular homologues have tyrosine-specific pro-
tein kinase activity, and it has been proposed that
phosphorylation at tyrosine of specific substrates may be
35 important in the transformation process. Treatment of

human carcinoma A431 cells with various TGFs derived from conditioned media of virally-transformed cells or human tumor cell lines (TGF- α) resulted in phosphorylation of tyrosine residues in the 160 K EGF receptor. The pattern of phosphorylation, however, was indistinguishable from that induced by EGF itself, and thus would not appear to be transformation-specific. Likewise, dissolution of actin fibers of Rat-1 cells occurs when they are treated with either TGF or EGF. It is clear that further research is needed to establish the relationships of the TGFs to the retrovirus transforming gene products and the mode of action of the TGFs in neoplastic transformation.

The following are summaries of examples which illustrate various aspects of this invention:

Example 1.

HPLC separation of TGF- α and TGF- β of MSV-transformed 3T3 cells. The acid/ethanol extract of MSV-transformed cells was chromatographed on Bio-Gel P-30 in 1 M acetic acid. The 7-10,000 MW TGF fraction was further purified on a μ Bondapak C18 column using a gradient of acetonitrile in 0.1% trifluoroacetic acid. Aliquots were assayed for colony-forming activity in the soft agar assay; in the presence of 2 ng/ml EGF; and in competition with 125 I-EGF in a radio-receptor assay.

Example 2.

HPLC purification on μ Bondapak CN columns of TGF- β from MSV-transformed mouse 3T3 cells and bovine salivary gland using a gradient of n-propanol in 0.1% trifluoroacetic acid. TGF- β of acid/ethanol extracts was purified on Bio-Gel P-30 and μ Bondapak C18 columns and then applied to CN columns. Aliquots were assayed for induction of colony growth of NRK cells in soft agar in the presence of 2 ng/ml EGF.

Example 3.

Synergistic interaction (activation) of TGF- β with TGF- α to induce the formation of large colonies of NRK cells in soft agar. Soft agar colony-forming activity of varying concentrations of μ Bondapak CN-purified TGF- β derived from MSV-transformed 3T3 cells was assayed either alone or in the presence of either CN-purified TGF- α derived from the same cells or murine EGF. Soft agar colony-forming activity of varying concentrations of EGF or TGF- α was assayed either alone or in the presence of TGF- β .

In Vivo Demonstration Of Wound Healing

After the above in vitro demonstrations of the operability of the compositions of this invention, it was considered critical to confirm that the compositions could work in clinical applications. For this purpose, TGFs were isolated on a relatively large scale from bovine sources and the wound healing activity of the compositions according to this invention were satisfactorily demonstrated using an experimental rodent wound healing protocol.

The examples which follow demonstrate not only that the compositions according to this invention are effective in vivo, but also that TGFs may be employed cross-species.

5 Example 4.

Purification and separation of TGF- α and TGF- β .

Bovine tissues, obtained fresh from the slaughterhouse and frozen immediately on dry ice, were extracted in 2 kg batches with acid/ethanol in accordance with
10 A. B. Roberts, et al., Proc. Natl. Acad. Sci., U.S.A.,
 77:3494 (1980). Extracts from 6-8 kg tissue were combined and chromatographed on Bio-Gel P-30 with 1 M acetic acid, using an 80 liter bed volume column.
15 The TGFs of extracts of bovine kidney or bovine salivary gland eluted in a broad peak between the RNase (13,700) and insulin (5,700) markers, as had been observed for the TGFs of mouse kidney and mouse salivary gland. TGFs at this stage of purification had a
20 specific activity approximately 10 to 25-fold higher than the acid/ethanol extracts, with a range of recovery of 150,000-200,000 colony-forming units per
25 kg tissue. Most of the in vivo studies reported below were done with salivary gland or kidney TGFs purified to this stage. The TGFs activity in vitro was enhanced approximately 20-fold by the presence of 2-5 ng EGF per ml in the assay, in accordance with this invention.

Following chromatography on Bio-Gel P-30, the bovine TGF- β were purified further by High Pressure Liquid Chromatography (HPLC) on μ Bondapak C18 columns using an acetonitrile gradient in 0.1 percent trifluoroacetic acid, followed by a second HPLC step on μ Bondapak CN columns using a gradient of n-propanol in 0.1 percent trifluoroacetic acid. After the two HPLC steps, analysis of the bovine TGF- β s from both salivary gland and kidney by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions showed a single band with an apparent molecular weight of 13,000 daltons. At this stage of purification, each of the bovine TGF- β 's had an absolute requirement for EGF for colony-forming activity. The yield of HPLC-purified TGF- β was approximately 20-100 μ g per kg tissue, with a total activity of 7,000-18,000 colony-forming units.

Example 5.

Wound healing protocol. In vivo activity of isolated salivary gland TGF- β and kidney TGF- β was measured in accordance with the protocol described by T. K. Hunt, et al., Amer. J. Surgery, 114:302 (1967). Six empty Schilling-Hunt wire mesh wound chambers were surgically inserted subcutaneously in the backs of rats, in paired symmetrical fashion (A-D, B-E, C-F) as shown below:

TABLE 2

head

A	D
B	E
C	F

tail

The rats respond to these chambers as if they were wounds, and eventually the chambers become filled with fibroblasts and collagen. By the fourth day after insertion, the chambers become encapsulated with connective tissue, but there are few cells within the chambers themselves. There is thus a defined, enclosed space within the chambers, where a wound healing response can be quantitatively measured. At this time, daily injections of TGF- β (0.1 ml, in sterile phosphate-buffered saline) into chambers A, B, and C were begun. To activate TGF- β activity, a low level of murine EGF was included in all TGF- β injections, unless noted otherwise. Chambers D, E, and F were used as controls, and were injected with either an amount of bovine serum albumin (BSA) alone or in combination with either TGF- β or EGF, such that the total protein was equivalent to the amount of TGF- β injected into chambers A, B, and C. Injections were made once daily for either 5 days (Table 2) or 9 days (Table 3). All injected materials were sterile. The rats were sacrificed 6 hours after the last TGF- β injection; in Table 3 they were injected with 0.5 mCi of thymidine- ^3H , specific activity 6.7 Ci/millimole (i.p.) together with the last TGF- β injection. The chambers were removed from the rats, all connective tissue on the outside of the wire mesh was peeled away, and then the contents of each chamber were determined.

Table 3. Wound healing response to bovine salivary gland or kidney TGF after 5 days of treatment. TGF- β s were prepared and injected as described. Each dose contained 25 times the amount of TGF- β found optimal for colony formation by NRK cells in a standard soft agar assay, and ranged from 18-42 colony forming units per dose. The amounts of protein injected per dose were: 7 μ g in Expts. 1, 4 and 5; 25 μ g in Expt. 3, and 0.7 μ g in Expt. 2. All doses of EGF were 20 ng. Total protein in wound chambers was measured by the method of Lowry, et al. Statistical analysis of the data was made by comparison of matched pairs of the chambers (A vs. D, B vs. E, C vs. F) shown in Table 2.

Expt.	Chamber A,B,C treatment	Chamber D,E,F treatment	Number of matched pairs of chambers	Milligrams of protein per cham- ber, average (15) A,B,C	D,E,F	Average ra- tio \pm standard error of mean ^a	It
1	TGF- β (Salivary P-30) + EGF	BSA	36	10	3.9	3.8 \pm 0.6	<0.001
2	TGF- β (Salivary HPLC) + EGF	BSA	9	8.4	2.9	4.6 \pm 1.0	<0.02
3	TGF- β (Kidney P-30) + EGF	BSA	9	8.1	3.5	5.2 \pm 1.5	<0.005
4	TGF- β (Salivary P-30) + EGF	EGF	9	9.6	5.3	2.1 \pm 0.3	<0.02
5	TGF- β (Salivary P-30) + EGF	TGF- β	9	11.2	9.6	1.4 \pm 0.3	0.5

^aAverage of each matched pair ratio, A/D, B/E, C/F
tOne sided \bar{p} values based on the sign test

^aJ. Biol. Chem., 193:265 (1951)

Table 4. Wound healing response to bovine salivary gland TGF- β after 9 days of treatment. Chambers A, B, and C were dosed once daily with 7 μ g of TGF- β (P-30) plus 20 ng of EGF. Chambers D, E and F were dosed with an equal amount of BSA.

Measurement	Number of matched pairs of chambers	Average content per chamber A,B,C	Average content per chamber D,E,F	Average t -test to standard error of mean	P
Protein, milligrams	30	24	15	1.6 \pm 0.05	<0.001
DNA, micrograms	30	21	8.6	2.6 \pm 0.16	<0.001
Thymidine- 3 H, cpm per microgram of DNA	30	45	30	1.7 \pm 0.09	<0.001
Collagen, milligrams	9	5.2	3.2	1.8 \pm 0.2	<0.005

Average of each matched ratio pair, A/D, B/E, C/F
One sided P values based on the sign test

Table 2 shows that 5 days of treatment of rats with TGF- β from either bovine salivary gland or bovine kidney caused a significant increase in total protein in the treated chambers, as compared to control chambers treated with an equivalent amount of bovine serum albumin (Experiments 1, 3). The salivary gland TGF- β was still highly active after two steps of purification by the high pressure liquid chromatography (Experiment 2). The effects observed are not the sole result of the minute amounts of EGF which had been used to potentiate the activity of TGF- β , since a highly significant difference between treated chambers A, B and C, compared to control chambers D, E and F was still observed when EGF was used as the control substance (Experiment 4). Furthermore, when all chambers were treated with TGF- β , and only A, B, and C were treated with EGF, no significant difference was observed (Experiment 5). At the end of Experiments 1-4, it was consistently observed that chambers A, B and C were more firmly fixed in the surrounding connective tissue than the respective matched control chambers, suggesting that effects of the TGF- β also were manifested in the area immediately surrounding the chambers.

In order to measure the effects of bovine salivary TGF- β on DNA and collagen content of the chambers, it was necessary to treat the animals for longer than 5 days. Table 3 shows the results of a larger experiment in which 13 rats were treated for 9 days. The increases in total protein, total DNA, thymidine incorporation into DNA, and total collagen were all highly sufficient. Histological examination of the contents of the chambers treated with TGF- β confirmed the occurrence of fibroblastic proliferation and formation of collagen. A

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sterile infiltrate of inflammatory cells was also found within both treated and control chambers.

The results obtained in both experiments indicate that TGF- β s when activated in accordance with this invention, can significantly accelerate a wound healing response.

Clinical Use Of The Compositions Of This Invention

The compositions of this invention, whose active ingredients are TGF- β activated by at least one of a TGF- α and an EGF, can reasonably be expected to have clinical use in the treatment of animals, particularly mammals, most particularly human beings. There are several sound bases for this conclusion.

It has been shown above, that in in vitro tests, the compositions can markedly increase the growth of cells without changing their genotype. An important characteristic of the components of the compositions of this invention, is that they do not appear to be species specific. That is, TGF- β from one species can be activated by TGF- α and/or EGF from other species. The cells whose growth is promoted can be of any type such as fibroblast or

epithelial, although it is considered that the growth promotion of fibroblast cells will have the greatest medical utility.

5 The in vivo experimental protocol disclosed above, with its very favorable results, clearly indicates that the compositions of this invention have utility in the treatment of traumata by the rapid promotion of the proliferation of the cells surrounding the traumata.

10 Two types of application of the compositions of this invention are contemplated.

15 The first, and preferred, application is topically for the promotion of surface wound healing. There are no limitations as to the type of wound or other traumata that can be treated, and these include (but are not limited to): first, second and third degree burns (especially second and third degree); surgical incisions, including those of cosmetic surgery; wounds, including lacerations, incisions, and penetrations; and
20 surface ulcers including decubital (bed-sores), diabetic, dental, haemophiliac, and varicose. Although the primary concern is the healing of major wounds by fibroblast cell regeneration, it is contemplated that the compositions may also be useful for minor wounds,
25 and for cosmetic regeneration of cells such as epithelial. It is also contemplated that the compositions may be utilized by the topical application to internal surgical incisions.

30 When applied topically, the compositions may be combined with other ingredients, such as carriers and/or adjuvants. There are no limitations on the nature of such other ingredients, except that they must be pharmaceutically acceptable, efficacious for their

intended administration, and cannot degrade the activity of the active ingredients of the compositions. When the compositions of this invention are applied to burns, they may be in the form of an irrigant, preferably in combination with physiological saline solution. The compositions can also be in the form of ointments or suspensions, preferably in combination with purified collagen. The compositions also may be impregnated into transdermal patches, plasters, and bandages, preferably in a liquid or semi-liquid form.

The second application is systemically for the healing of internal wounds and similar traumata. Such an application is useful provided that there are no, or limited, undesirable side-effects, such as the stimulation of neoplastic cellular growth.

When applied systemically, the compositions may be formulated as liquids, pills, tablets, lozenges, or the like, for enteral administration, or in liquid form for parenteral injection. The active ingredients may be combined with other ingredients such as carriers and/or adjuvants. There are no limitations on the nature of such other ingredients, except that they must be pharmaceutically acceptable, efficacious for their intended administration, and cannot degrade the activity of the active ingredients of the compositions.

The amount of activating agent (TGF- α s or EGFs) present depends directly upon the amount of TGF- β s present in the activated compositions of this invention. There are indications that the activation is not catalytic in nature, and that therefore approximately stoichiometric (equimolar) quantities are preferred.

The amount of activated composition to be used in the methods of this invention cannot be stated because of the nature of the activity of TGFs and the nature of healing wounds and/or other traumata. As indicated above, the TGFs activate cells by binding to receptor sites on the cells, after which the TGFs are absorbed and utilized by the cells for the synthesis of new protein, resulting in cell multiplication. Thus, the TGFs are consumed by the cell regenerating process itself, rather than acting in an enzymatic or other catalytic manner. Receptors for EGFs have been found on a wide variety of fibroblastic, epithelial, and parietal cells, as disclosed in Gonzalez, et al., J. Cell. Biol., 88:108-144 (1981). Further, it has been calculated that there are 3,000 EGF binding (receptor) sites for each rat intestinal epithelial cell, as disclosed in M. E. Lafitte, et al., FEBS Lett., 114(2):243-246 (1980). It must also be obvious that the amount of a cell growth promoting substance (such as the compositions of this invention) that must be utilized will vary with the size of the wound or other traumata to be treated.

Since the compositions of this invention both provoke and sustain cellular regeneration, a continual application or periodic reapplication of the compositions is indicated.

The amount of active ingredient per unit volume of combined medication for administration is also very difficult to specify, because it depends upon the amount of active ingredients that are afforded directly to the regenerating cells of the wound or other traumata situs. However, it can generally be stated that the TGF- β s should preferably be present in an amount of at least about 1.0 nanogram per milliliter of combined composition, more preferably in an amount up to about 1.0 milligram per milliliter.

Additional Embodiments Utilizing the Compositions
of this Invention

5 In addition to utilizing the activated TGF- β compositions of this invention by themselves, it is possible to use them in combination with secondary growth factors.

10 The activated transforming growth factors of this invention may be physically admixed with one or more of many other (secondary) peptide and non-peptide growth factors. Such admixtures may be administered in the same manner and for the same purposes as the activated transforming growth factors of this invention utilized alone, to enhance their activity in promoting cell proliferation and repair.

15 The useful proportions of activated transforming growth factor to secondary growth factors are 1:1-10 mols, with about equimolar amounts being preferred.

20 The secondary growth factors may be used alone or in any physiologically and pharmaceutically compatible combination.

The known secondary growth factors, in approximately descending order of usefulness in this invention (by group), include:

- 25
1. platelet-derived growth factors
 2. fibroblast growth factors
 angiogenesis factors
 3. insulin-like growth factors including
 somatomedins

4. insulin
nerve growth factors

5. anabolic steroids.

5 In addition to the above known secondary growth factors, it is reasonable to expect that as yet undiscovered secondary growth factors will be useful in admixture.

10 This invention also incorporates the inactive intermediate substance TGF- β per se. Prior to this invention, this substance had not been isolated or identified. TGF- β is believed to be substantially the same or very similar for each animal species, regardless of the individual of that species or the particular body cells from which it is derived.

15 Since TGF- β has been shown to be non-species-specific between rodents, cattle, and human beings, it is also reasonable to believe that the substance is substantially the same or very similar when derived from any mammal, and possibly from any animal source.

20 It should be noted, moreover, that this invention includes TGF- β regardless of the source from which it is isolated or derived, including genetically engineered cells. It is well within the capabilities of biochemical technology to genetically engineer

25 a cell to produce TGF- β at the present time.

Administration of Unactivated TGF- β

It is believed that TGF- β has no wound-healing or other tissue-repair activity unless it has been activated by an agent as described above.

5 However, it is noted that Table 3 Experiment 5, supra, appears to indicate statistically similar results for TGF- β activated with EGF (chambers A, B, C) and TGF- β per se (chambers D, E, F). The most logical explanation for this, is that
10 the TGF- β per se was activated by a TGF already present in the test animal. Various TGFs, such as EGF, are known to be present in blood plasma.

 Thus, the results of Experiment 5 are not inconsistent with this invention, but instead
15 constitute a variant embodiment thereof. Specifically, TGF- β per se may be administered, in accordance with this invention, instead of activated TGF- β , when there are sufficient endogenous activating agents present in an animal,
20 to activate an amount of TGF- β sufficient to promote cell proliferation and tissue repair. It is anticipated that in an animal suffering from the traumata contemplated herein, there usually will not be sufficient endogenous activating
25 agents present.

 The disclosures of the following applications, which were filed on the same date as the present continuation-in-part application, are entirely incorporated herein by reference:

1. "Transforming Growth Factor-beta From Human Platelets", by Richard K. Assoian, Charles A. Frolik, Michael B. Sporn, and Anita B. Roberts.
2. "Transforming Growth Factor-beta From Human Placentas", by Charles A. Frolik, Richard K. Assoian, Michael B. Sporn, and Anita B. Roberts.

WHAT IS CLAIMED IS:

1. A composition for the promotion of cell proliferation and tissue repair in animals whose active ingredients consist essentially of:

- beta-type transforming growth factors (TGF- β); and
- at least one activating agent present in an amount sufficient to activate said TGF- β , selected from the group consisting of epidermal growth factors (EGF) and alpha-type transforming growth factors (TGF- α),
- which active ingredients are present in an amount sufficient to promote cell proliferation.

2. The composition of Claim 1, used for the topical promotion of fibroblast cell proliferation in mammals wherein the TGF- β and at least one activating agent are present in approximately equimolar amounts.

3. The composition of Claim 2, wherein said TGF- β is present in an amount of at least 1.0 nanogram per milliliter of composition.

4. The composition of Claim 3, wherein said TGF- β is present in an amount up to 1 milligram per milliliter of composition.

5. The composition of Claim 2, wherein the activating agent is an EGF.

6. The composition of Claim 2, wherein the activating agent is a TGF- α .

7. The composition of Claim 2 further containing at least one pharmaceutically acceptable carrier.

8. The composition of Claim 7, wherein the carrier is a purified collagen and the composition is a suspension for topical application.

9. The composition of Claim 7, wherein the carrier is a physiological saline solution and the composition is an irrigant.

10. The composition of Claim 1, used for the systemic promotion of cell proliferation and tissue repair in mammals wherein the TGF- β and at least one activating agent are present in approximately equimolar amounts and wherein the composition further contains a pharmaceutically acceptable carrier.

11. A method of treating mammalian topical wounds, burns, ulcers or other traumata comprising at least one topical application of the composition of Claim 2 in a pharmaceutically acceptable carrier.

decubitus ulcers, and many other conditions, at present there is no practical way to accelerate wound healing with pharmacological agents. Although it is suggested in Tissue Growth Factors, supra, at p. 123 that EGF
5 might be of benefit in this area, it has yet to be extensively used in a practical way for wound healing.

SUMMARY OF THE INVENTION

This invention affords compositions for the promotion of cell proliferation in animals, especially
10 fibroblast cells in human beings. The compositions have as their active ingredients, beta-type transforming growth factor (TGF- β) and an activating agent. The activating agents of this invention are selected from at least one of epidermal growth factor (EGF) and
15 alpha-type transforming growth factor (TGF- α).

The TGF- β and the activating agent are preferably present in about equimolar amounts, and the active ingredients are present in an amount at least sufficient to stimulate cell proliferation (tissue repair).

20 As another embodiment, the activated TGF- β compositions of this invention may be admixed with other (secondary) growth factors to enhance their activity.

The compositions may be formulated in any suitable carrier for topical application, such as physiological
25 saline solution or purified collagen suspension. The compositions also may be formulated in any suitable carrier for systemic administration.

The method of topical administration of the compositions of this invention is by direct application to
30 a burn, wound, or other traumata situs. Periodic or continual further administration may be preferably indicated in most instances, since the active ingredients are physiologically utilized by the cells whose growth is being stimulated.

As a further embodiment, the compositions of this invention may be administered systemically by injection, enterally, transdermal patches, and the like, depending upon the nature and site of the traumata to be treated.

5

DETAILED DESCRIPTION OF THE INVENTION

10 The term, "transforming growth factor" (TGF) has been defined to include the set of polypeptides which confer the transformed phenotype on untransformed indicator cells. The transformed phenotype is operationally defined by the loss of density-dependent inhibition of growth in monolayer, overgrowth in monolayer, characteristic change in cellular morphology, and acquisition of anchorage independence, with the resultant ability to grow in soft agar. Untransformed, 15 non-neoplastic cells will not form progressively growing colonies in soft agar, while the property of anchorage-independent growth of cells in culture has a particularly high correlation with neoplastic growth in vivo.

20

Although TGFs were first discovered in the conditioned medium of virally-transformed neoplastic mouse cells, the application of the acid/ethanol method for extraction of peptides from tissues has now shown that TGFs can be found in almost all tissues, both neoplastic and non-neoplastic, from all species of animals 25 that have been examined thus far. Although TGF activity is usually measured with an in vitro phenotypic transformation assay, this does not imply that TGF

28. A transforming growth factor having an amino acid composition consisting essentially of the following residues per mole based upon an apparent molecular weight of 25,000: (wherein ND means not determined)

Aspartic acid	25 \pm 1
Threonine	7 \pm 1
Serine	16 \pm 2
Glutamic acid	26 \pm 1
Proline	N.D.
Glycine	15 \pm 3
Alanine	18 \pm 1
Half-cystine	16 \pm 2
Valine	17 \pm 1
Methionine	3 \pm 0
Isoleucine	11 \pm 1
Leucine	25 \pm 1
Tyrosine	17 \pm 2
Phenylalanine	8 \pm 1
Histidine	8 \pm 1
Lysine	22 \pm 2
Tryptophan	N.D.
Arginine	12 \pm 1

29. A transforming growth factor having a partial amino acid sequence as determined by Edman Degredation consisting essentially of the following for each of its two subunits:

Ala-Leu-Asp-Thr-⁵Asn-Tyr-CMC-Phe-Ser-¹⁰Ser-Thr-Glu-Lys-¹⁵Asn-CMC-,

where CMC is Half-cystine or cysteine, determined as S-carboxymethylcysteine.

30. A method of treating mammalian topical wounds, burns, ulcers or other traumata comprising at least one topical application of the composition of Claim 23 in a pharmaceutically acceptable carrier.

31. A method of treating mammalian systemic traumata comprising at least one systemic administration of the composition of Claim 23 in a pharmaceutically acceptable carrier.

